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2 **Gastroprotective activity of methanol leaves extract of *Barleria prionitis***
3 **Linn. on ethanol and indomethacin induced ulcer in rats**

4 Manjusha¹, Vipin Kumar*¹, Surender Singh²

5 ¹Institute of Pharmaceutical Sciences, Kurukshetra University,
6 Kurukshetra-136119, India

7 ²All India Institute of Medical Sciences, New Delhi- India

8 vipbhardwaj@rediffmail.com

9 **Abstract**

10 **Aim:** *Barleria prionitis* L. (Family Acanthaceae) is a medicinal plant found road side in India
11 and whole plant or its various parts like leaves, root, bark, stem and flowers are used
12 traditionally for various treatments like toothache, inflammation, boils, glandular swellings
13 and ulcer. Leaf juice is useful in gastric ulcer. Here, we attempt to prove the use of this plant
14 in gastroprotection.

15 **Study design:** This study was conducted to evaluate the antiulcer activity of methanol extract
16 obtained from the leaves of *Barleria prionitis* Linn.

17 **Place and duration of the study:** The experiments were conducted at pharmacology lab of
18 Institute of Pharmaceutical Sciences, Kurukshetra University during the period of July 2012
19 to December 2012.

20 **Material and methods:** Antiulcer activity was performed using the protocols of ulcer
21 induced by ethanol and indomethacin at two different doses (250 and 500mg/kg). Parameters
22 like volume of gastric juice, pH, free acidity, total acidity, aspartate amino transferase (AST)
23 and alanine amino transferase (ALT) were also determined in ethanol induced ulcer model.

24 **Results:** The reduction in ulcer index in *Barleria prionitis* treated animals was found to be
25 statistically significant (P=.05), when compared with control groups in both the models.
26 Significant changes were observed in total acidity at dose 500mg/kg only and changes were
27 significant in AST, ALT levels at both the doses. Other parameters showed non-significant
28 results.

29 **Conclusion:** The results of the present study show that the methanols extract of *Barleria*
30 *prionitis* possess antiulcer activity. This work supports the traditional use of this plant in
31 treating gastric ulcer.

32
33 **Keywords:** *Barleria prionitis*, Gastroprotective activity, Ulcer index, Methanol extract,
34 Ethanol

36

37 **1. Introduction**

38

39 Gastric hyperacidity is a very common global problem that affects millions of people
40 worldwide [1,2]. In gastric ulcer their occurs imbalance between aggressive (acid-pepsin
41 secretions) and protective factors (such as mucus secretion, mucosal barrier, cell
42 regeneration, blood flow and prostaglandins) [3,4]. The current treatment of peptic ulcer is
43 mainly done with H₂ receptor antagonists, proton pump inhibitors, and antimuscarinics. But,
44 most of these treatments produce adverse reaction like, hypersensitivity, arrhythmia,
45 impotence, gynecomastia and hematopoietic disorders [5,6,7,8]. Therefore, there is
46 requirement for new and safer treatment, with fewer side effects. Plants extracts are among
47 the suitable treatments for the prevention of gastric ulcer [9].

48 *Barleria prionitis* L. (Family Acanthaceae; commonly known as Vajradanti) is a medicinal
49 plant found throughout South Africa, India, Sri-Lanka, and tropical Asia [10,11]. In India
50 whole plant or its various parts like leaves, root, bark, stem and flowers are used traditionally
51 for various treatments like toothache, inflammation, boils, glandular swellings, whooping
52 cough etc [12,13,14,15]. The leaf juice is useful in stomach problems, ulcer, fever and urinary
53 affections [16]. The whole plant including roots is used to induce diuresis. Plant is also useful
54 in jaundice, hepatic problems and dropsy [17,18].

55 Phytochemical studies on hydro-methanolic extract of *B. prionitis* showed the presence of
56 glycosides, steroids, tannins and flavonoids [19]. Iridoid glycosides, shanzhiside methyl ester,
57 6-O-trans-p-coumaroyl-8-O-acetylshanzhiside methyl ester, barlerin, acetylbarlerin, 7-
58 methoxydideroside and lupulinoside have been isolated from aerial parts [20]. None study
59 was conducted scientifically to prove the gastroprotective effect of *B. prionitis* leaves. Hence
60 the present study was conducted to evaluate the antiulcer properties of methanolic extract of
61 *B. prionitis* Linn.

62

63 **2. Materials and methods**

64

65 **Plant material**

66 The leaves of *Barleria prionitis* were collected from Ashoka nursery Gharunda, Karnal,
67 Haryana, India in the month of March, 2011. Then, collected leaves were positively identified
68 by Dr. H.B. Singh, Head, Raw Materials Herbarium and Museum (RHMD), New Delhi. A
69 voucher specimen of the plant (Ref. No. NISCAIR/RHMD/CONSULT/-2010-11/1497/95)
70 has been preserved there for future references.

71

72 **2.1. Extraction**

73

74 The leaves were thoroughly washed under running tap water so as to remove any type of
75 contamination. Then washed leaves were air dried in shade, powdered in grinder and passed
76 through sieve of mesh size no-40. The dried powder was first defatted by petroleum ether and
77 then successive extraction was done with chloroform and methanol by hot Soxhlet extraction
78 method. The methanol extract was concentrated in a rotary evaporator under reduced
79 pressure. The dried crude extract was collected and preserved in airtight glass container at
80 4°C - 8°C.

81

82 **2.2. Preliminary phytochemical studies**

83

84 To determine the chemical constituents, the methanol extract obtained was thus subjected to
85 phytochemical analysis [21].

86

87 **2.3. Antiulcer activity**

88

89 ***2.3.1. Experimental animals***

90 Healthy Wistar rats of either sex were obtained from a disease free animal house of
91 Chaudhary Charan Singh, Haryana Agriculture University, Hisar, Haryana (India). The
92 animals were housed in the animal house, Institute of Pharmaceutical Sciences, Kurukshetra
93 University, Kurukshetra, Haryana (India). Rats were fed with commercially available feed
94 and were maintained under standard conditions of temperature ($25^{\circ}\text{C} \pm 5^{\circ}\text{C}$), relative
95 humidity ($55 \pm 10\%$), and 12/12 h light/dark cycle. They were housed in standard
96 polycarbonate cages with wire mesh top and husk bedding.

97 ***2.3.2. Experimental design***

98 Wistar rats weighing between 175-250 g of either sex were used for antiulcer study. All
99 animals were divided into 4 groups of 6 animals. Before the experiments, animals were
100 deprived of food but allowed free access to water.

101

102 ***2.3.3. Dose and route of administration***

103 For experimentation 250mg/kg and 500mg/kg doses of BPM were used. Fresh drug solutions
104 were prepared in sterile distilled water at the time of administration and were administered
105 Per Oral (p.o.) so as to avoid any additional stress to the animals.

106

107 **2.3.4. Group designing for ethanol and indomethacin induced ulcer models**

108 Group I (Control): Animals received only distilled water; Group II (BPM 250): Animals
109 received *B. prionitis* (250mg/kg, p.o.) I hr before the ulcerogenic procedure; Group III (BPM
110 500): Animals received *B. prionitis* (500mg/kg, p.o.) I hr before the ulcerogenic procedure;
111 Group IV (Standard): Animals received ranitidine (50mg/kg, p.o.) I hr before the ulcerogenic
112 procedure.

113 **2.3.5. Ethanol induced gastric mucosal lesions**

114 This is a widely used model that seems to cause gastric ulcer. The activity was performed
115 according to the slightly modified method of Mizui and Dotuchi [22]. Rats were fasted for 36
116 h before administration of absolute ethanol (1.0mL). The group I was given only distilled
117 water. The extract (250 mg/kg, 500mg/kg, p.o.) and ranitidine (50mg/kg, p.o.) as standard
118 drug, were given to Group II, III and IV respectively. One hour after treatment, all the rats
119 received ethanol to induce gastric ulcer. Another one hour later, animals were sacrificed by
120 cervical dislocation. The stomachs were removed, cut and opened along the greater curvature,
121 washed with normal saline to remove the gastric contents and observed for the severity of the
122 ulcers. The pH and volume of gastric juice was measured after centrifugation at 2000rpm for
123 10 min. From the supernatant, aliquots were taken for the determination of total and free
124 acidity. The percentage protection was calculated using the following formula:-

$$125 \% I = (UI \text{ of control} - UI \text{ of test}) \times 100 / UI \text{ of the control}$$

126 Where I = Inhibition, UI= Ulcer index

127 **2.3.6. Ulcer indexing**

128 The mucosal layer of the stomach was observed under a magnifying lens and ulcers were
129 checked. The area (mm²) of all lesions was measured using digital callipers' to give a gastric
130 damage score. The ulcer index was determined using the following formula [23].

$$131 UI = 10/X$$

132 Where X= total mucosal area/total ulcerated area

133 **2.3.7. Total acidity and free acidity determination**

134 1.00mL of centrifuged and filtered gastric juice was taken in a conical flask. Two drops of
135 1% phenolphthalein indicator for total acidity and Topfer's reagent for free acidity was added
136 to it. It was titrated against 0.1mol/L sodium hydroxide until a permanent pink color (total
137 acidity) or canary yellow colour (free acidity) was observed. The total/free acidity is
138 expressed as meq./L by the following formula:-

$$139 \text{ Total/free acidity} = n \times 0.01 \times 36.45 \times 1000$$

140 Where, n is the volume of NaOH consumed, 0.01 is normality of NaOH, 36.45 is molecular
141 weight of NaOH, 1000 is the factor (to be represented in litre).

142 **2.3.8. Indomethacin induced gastric ulcers**

143 In this model, the gastric lesions are induced by the inhibition of prostaglandin synthesis.
144 Activity was performed according to method of Djahanguiri [24] and 24 h fasted rats were
145 used for study. Group I animals were treated orally with distilled water. The extract (250
146 mg/kg, 500mg/kg, *p.o.*) and ranitidine (50mg/kg, *p.o.*) as standard drug, were given to Group
147 II, III and IV respectively. One hr. after the treatment, 20mg/kg of indomethacin (dissolved in
148 2% sodium bicarbonate) was administered orally. After 4 h, all animals were sacrificed by
149 cervical dislocation. The stomachs were isolated, washed with normal saline and various
150 parameters like ulcer index, free acidity and total acidity were measured as discussed above
151 [25].

152 **2.3.9. Serum biochemical parameters**

153 Blood samples were analysed for AST and ALT level estimation in ethanol induced gastric
154 lesions.

155 **2.4. Statistical analysis**

156 All the values were expressed as mean±standard error of mean. The statistical significance of
157 difference among groups was analysed using one-way ANOVA. A value of $P < 0.05$ was
158 considered significant.

159

160 **3. Results**

161 **3.1. Preliminary phytochemical screening**

162 The percentage yield of petroleum ether, chloroform and methanol leaf extracts were found to
163 be 4.9, 6.9 and 16.7% (in weight). Preliminary phytochemical screening of methanol extract
164 showed the presence of steroids, alkaloids, saponins, glycosides and flavonoids.

165

166 **3.2. Antiulcer activity**

167 **3.2.1. Ethanol induced gastric ulcer**

168 In this study, BPM was screened for gastroprotective activity against ethanol induced gastric
 169 ulcer in rats. The absolute ethanol administration (*p.o.*) induced severe ulceration. BPM and
 170 ranitidine groups showed the significant reduction in incidence and severity of ulceration.
 171 BPM and ranitidine showed a significant change in ulcer index when compared with the
 172 control group $P<0.01$ (Table 1). BPM and ranitidine showed slight changes in pH, volume of
 173 gastric juice, free acidity and total acidity but changes were not significant when compared
 174 with control group except total acidity in BPM (500mg/kg) treated group, $P<0.01$ (Table 2).

175

176 **Table 1. Ulcer index of ethanol and indomethacin induced gastric ulcers**

Model	Group	Dose(mg/kg body weight)	Ulcer index	% Inhibition
Ethanol	Ethanol	--	0.90±0.01	-
	BPM	250	0.43±0.02**	52.2%
	BPM	500	0.29±0.04**	67.7%
	Ranitidine	50	0.22±0.02**	75.5%
Indomethacin	Indomethacin	20	1.35±0.15	-
	BPM	250	0.51±0.03**	62.2%
	BPM	500	0.40±0.02**	70.3%
	Ranitidine	50	0.51±0.03**	62.2%

177 *Results as mean±S.E.M. for six rats. Statistical comparison was performed using ANOVA*
 178 *followed by the Dunnett's test. ** $P<0.01$ when compared with control group.*

179

180

181 **Table 2. Volume of gastric juice, pH, free acidity and total acidity in ethanol induced**
 182 **gastric ulcer**

Group	Dose(mg/kg)	Volume of gastric juice(ml)	pH	Free acidity(mmol/h)	Total acidity(mmol/h)
Ethanol	--	2.08±0.01	4.42±0.06	0.53±0.008	1.22±0.008
BPM	250	2.55±0.18	4.51±0.06	0.48±0.02	1.23±0.02
BPM	500	2.22±0.18	4.40±0.05	0.46±0.02	0.81±0.01**
Ranitidine	50	2.72±0.27	4.51±0.03	0.51±0.03	1.26±0.02

183 *Results as mean±S.E.M. for six rats. Statistical comparison was performed using ANOVA*
 184 *followed by the Dunnett's test. ** $P<0.01$ when compared with control group.*

185

186

187 **3.2.2. Biochemical parameters**

188 Ethanolic group induced ulcer showed a increase in liver enzymes (ALT and AST) as shown
 189 in (Table 3). When rats were pretreated with BPM (250mg/kg and 500mg/kg) and ranitidine,
 190 there were significant reductions in serum concentration of these markers, $P<0.01$, $P<0.05$.

191

192

193 **Table 3. Effect of BPM extract on liver functions tests in ethanol induced gastric ulcers**

Group	Dose(mg/kg)	ALT(IU/L)	AST(IU/L)
Ethanol	--	65.98±2.5	353±2.1
BPM	250	56.08±2.1*	325±7.1**
BPM	500	45.44±2.0**	305±4.8**
Rantidine	50	55.18±3.5*	331±3.07*

194 *Results as mean±S.E.M. for six rats. Statistical comparison was performed using ANOVA*
 195 *followed by the Dunnett's test. **P<0.01, *P<0.05 when compared with control group.*
 196

197 **3.3.3. Indomethacin induced mucosal lesions**

198 Indomethacin (20mg/kg, *p.o.*) administration induced severe gastric mucosal damage. BMP,
 199 at tested doses 250 and 500mg/kg, showed significant gastroprotective effect against gastric
 200 lesions, *P*<0.01. Standard drug ranitidine (50mg/kg, *p.o.*) included in the study as positive
 201 control also exhibited significant protection, *P*<0.01(Table1 and Figure 1).

202 **Figure1. Macroscopical view of rat stomach in indomethacin induced gastric**
 203 **ulcer**



204 Ethanol Group



205 Ranitidine Group

204

205

206



207

208

209 BPM (250mg/kg)

BPM (500mg/kg)

210 **4. Discussion**

211 Various noxious stimuli of endogenous (acid and pepsin) and exogenous (drugs and alcohol)
212 origin, constantly comes in contact with gastric mucosa. Gastric mucosal defensive
213 mechanism, like mucosal blood flow, bicarbonate and mucus secretion protect the gastric
214 mucosa from damage. It is generally believed that it results from an imbalance between
215 aggressive factor (acid, pepsin) and defensive factors (mucous secretions, prostaglandins)
216 [26]. Therapeutic agents including different plant extracts are used to regain the balance by
217 inhibiting the gastric acid secretion or by increasing the mucous production, stabilizing the
218 surface epithelial cells. Herbs are the one of the most promising sources of new drugs as
219 these are free of or having very less side effects and adverse reactions.

220 The methanol extract of *B. prionitis* was used to evaluate gastro protective activity by using
221 ethanol and indomethacin induced gastric ulcers. Ethanol is considered very important in
222 inducing gastric ulcers. Ethanol induced ulcer depends upon lots of factors. One reason may
223 be its rapid penetration into gastric mucosa, which may cause more mucosal permeability and
224 release of vasoactive factors that leads to gastric damage [27,28]. The other factor responsible
225 may be formation of reactive oxygen species, which cause an imbalance between oxidant and
226 antioxidant process, that results rupture of blood vessels, thus contributes to the haemorrhage,
227 tissue necrosis and disrupting the protective mucosal barrier [29,30]. Indomethacin is
228 considered to induce ulcers in the stomach mainly due to inhibition of prostaglandin synthesis
229 [31]. To study the side effects of *B. prionitis* on liver, serum AST and ALT were determined
230 in ethanol induced gastric ulcer model. Control group animals showed, increase of serum
231 concentration of these enzymes that indicates hepatic injury since level of these enzymes
232 increases in chemically triggered tissue injury. *B. prionitis* administration decreased the
233 levels of AST and ALT that shows its tissue damage preventing action.

234 The preliminary phytochemical analysis indicated the presence of flavonoids, sterols,
235 glycosides, saponins. These secondary metabolite classes are related to gastro protective
236 activity. There are many studies related to the antiulcer genic properties of flavonoids
237 [32,33]. Leaves of the plant also contain saponins. Saponins exhibit ulcer protective effect by
238 selective inhibition of prostaglandin $F_{2\alpha}$ and by protection of gastric mucosa [29,34]. In view
239 of this fact it is suggested that gastro protection elucidated by the methanol extract of *B.*
240 *prionitis* may be related to the presence of these phytoconstituents.

241 **5. Conclusions**

242 The results provide support for the traditional use of this plant in the treatment of gastric
243 ulcer. However, the data so far obtained do not indicate the specific mechanism(s)
244 responsible for the antiulcer activity. Further studies are required to isolate the active
245 components and to elucidate their mechanism of action. In conclusion, the results show that
246 methanol extract of *Barleria prionitis* Linn. possess gastro protective activity.

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252 Competing interests

253 The authors declare that they have no competing interests.

254 Author's contribution

255 1st author wrote the protocol, performed the study and statistical analysis 2nd author designed
256 the study, managed the analysis, write the first draft of manuscript, 3rd author helped in the
257 literature study and finally all the authors read and approved the final manuscript.

258 Ethical Approval

259 All authors hereby declare that, Principles of laboratory animal care (NIH publication No.85-
260 23, revised 1985) were followed. All the experimental procedures and protocols used in the
261 study were reviewed by the Institutional Animal Ethics Committee (IAEC) (Register
262 Number: 562/GO/02/a/CPCSEA) and were in accordance with the CPCSEA guidelines,
263 Government of India.

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265 References

- 266 1. Jainu M, Devi CSS. Antiulcerogenic and ulcer healing effects of *Solanum nigrum*
267 (L.) on experimental ulcer models: possible mechanism for the inhibition of acid
268 formation. J Ethnopharmacol. 2006; 104: 156-163.
- 269 2. Klein-Junior LC, Gandolfi RB, Santin JR, Lemos M, Cechinel FV, Andrade SF.
270 Antiulcerogenic activity of extract, fractions, and some compounds obtained from
271 *Polygala cyparissias* St Hillaire & Moquin (Polygalaceae). Naunym-Schmiedberg's
272 Archives of Pharmacology. 2010; 381: 121-126.
- 273 3. Muralidharan P, Srikanth J. Antiulcer activity of *Morinda citrifolia* Linn fruit extract.
274 J Sci Res. 2009; 1(2): 345-352.
- 275 4. Lima ZP, Severi JA, Pellizzon CH, Brito ARMS, Solis PN, Caceres A et al. Can the
276 aqueous decoction of mango flowers be used as antiulcer agent? J Ethnopharmacol.
277 2006; 106: 29-37.
- 278 5. Chan FK, Leung WK. Peptic ulcer disease. Lancet. 2002; 360: 933-941.
- 279 6. Malfertheiner P, Chan FKL, McColl KEL. Lancet. 2009; 374: 1449-1461.
- 280 7. Sheen E, Triadafilopoulos G. Adverse effects of long- term proton pump inhibitor
281 therapy. Digestive Diseases and Sciences. 2011; 56: 931-950.

- 282 8. Bighetti AE, Antonio MA, Kohn LK, Rehder VLG, Foglio MA, Possenti A, et al.
283 Antiulcerogenic activity of a crude hydroalcoholic extract and coumarin isolated from
284 *Mikania laevigata* Schultz Bip. *Phytomedicine*. 2005; 12: 72-77.
- 285 9. Rodriguez A, Theoduloz C, Yanez T, Becerra J, Schmeda HG. Gastroprotective and
286 ulcer healing affect of ferruginol in mice and rats: assessment of its mechanism of
287 action *in vitro* models. *Life Sci*. 2006; 79: 2503-2509.
- 288 10. Chopra RN, Nayar SL, Chopra IC. In *Glossary of Indian Medicinal Plants*, CSIR:
289 New Delhi; 1965, p. 33.
- 290 11. Gupta HM, Saxena VK. A new acylated luteolin-7-O- β -D-glucoside from the roots of
291 *Barleria prionitis* (Linn.). *Nat Acad Sci Lett*. 1984; 7: 187-189.
- 292 12. Daniel M. *Medicinal Plants: Chemistry and Properties*. 1st ed. USA: Science
293 Publishers; 2006, p.78.
- 294 13. Aneja KR, Joshi R, Sharma C. Potency of *Barleria prionitis* L. bark extracts against
295 oral disease causing strains of bacteria and fungi of clinical origin. *New York Sci J*.
296 2010; 3: 5-12.
- 297 14. Banerjee D, Maji AK, Mahapatra S, Banerji P. *Barleria prionitis* Linn : A Review of
298 its Traditional Uses, Phytochemistry, Pharmacology and Toxicity. *Res J Phyto*. 2012;
299 6(2): 1-11.
- 300 15. Khare CP. *Indian Herbal remedies: Rational Western Therapy, Ayurvedic and Other*
301 *Traditional usage, Botany*. 1st ed. New York: Springer; 2004, p. 93-94.
- 302 16. Khare CP. *Indian Medicinal Plants: An Illustrated Dictionary*. 1st ed. New York:
303 Springer Science; 2007, p. 82-83.
- 304 17. Shukla P, Singh A, Gawri S, Alexende A, Sonwane S. *In vitro* propagation of
305 *Barleria prionitis* Linn and its antibacterial activity. *Int J Pharma Prof Res*. 2011; 2:
306 198-200.
- 307 18. Khadse CD, Kadke RB. Antiinflammatory activity of aqueous extract fractions of
308 *Barleria prionitis* L. roots. *Asian J Plant Sci Res*. 2011; 1: 63-68.
- 309 19. Maji AK, Bhadra S, Mahapatra S, Banerji P, Banerjee D. Mast cell stabilization and
310 membrane protection activity of *Barleria prionitis* L. *Pharmacog. J* 2011; 3: 67-71.
- 311 20. Ata A, Kalhari KS, Samarasekera R. Chemical constituents of *Barleria prionitis* and
312 their enzyme inhibitory and free radical scavenging activities. *Phytochem Lett* 2009;
313 2: 37-40.
- 314 21. Khandelwal KR. *Practical pharmacognosy techniques and experiments*. 3rd ed. Pune:
315 Nirali Prakashan; 1996, p. 171-172.
- 316 22. Mizui T, Doteuchi M. Effect of polyamines on acidified ethanol-induced gastric
317 lesion in rats. *The Jpn J Pharmacol*. 1983; 33: 939-945.
- 318 23. Ganguly AK. A method for quantitative assessment of experimentally produced
319 ulcers in the stomach of albino rats. *Experientia*. 1969; 25(11):1224.
- 320 24. Djahanguiri B. The production of acute gastric ulceration by indomethacin in the rat.
321 *Scandinavian J Gastroenterology*. 1969; 4: 265-267.
- 322 25. Dijoseph JF, Eash JR, Mir GN. Gastric antisecretory and antiulcer effects of
323 WHR1582A, a compound exerting alpha-2 adrenoceptor agonist activity. *J Pharmacol*
324 *Exp Ther*. 1987; 24(1): 97-102.

- 325 26. Piper DW, Stiel DD. Pathogenesis of chronic peptic ulcer, current thinking and
326 clinical implications. *Med Prog.* 1986; 2: 7-10.
- 327 27. Suleyman H, Buyukokuroglu ME, Koruk M, Akcay F, Kiziltunc A, Gepdiremen A.
328 The effects of *Hippophae rhamnoides* L. extract on ethanol-induced gastric lesion and
329 gastric tissue glutathione level in rats: A comparative study with melatonin and
330 omeprazole. *Indian J Pharmacol.* 2001; 33: 77-81.
- 331 28. Narayan S, Devi RS, Jainu M, Sabitha KE, Devi CSS. Protective effect of polyherbal
332 drug, ambrex in ethanol-induced gastric mucosal lesion in experimental rats. *Indian J*
333 *Pharmacol.* 2004; 36: 34-37.
- 334 29. Lewis DA, Hanson PJ. Antiulcer drugs of plant origin. *Prog Med Chem.* 1991; 28:
335 201-231.
- 336 30. Skiric P, Seiwerth S, Grabarevic Z, Rucman R, Petek M, Jagic V et al. The influence
337 of a novel pentadecapeptide, BPC 157, on N(G)-nitro-L-arginine methylester and L-
338 arginine effects on stomach mucosa integrity and blood pressure. *European Journal of*
339 *Pharmacol.* 1997; 332: 23-33.
- 340 31. Sagar V, Ahamed RN. Gastric mucosal cellular changes induced by indomethacin
341 (NSAID) in male albino rats. *Indian J Exp Bio.* 1999; 37(4): 365-369.
- 342 32. Gracioso JS, Vilegas W, Hiruma-Lima CA, Brito ARMS. Effects of tea from *Turnera*
343 *ulmifolia* L. on mouse gastric mucosa support the Turneraceae as a new source of
344 antiulcerogenic drugs. *Bio Pharma Bull.* 2002; 25: 487-491.
- 345 33. Gonzalez FG, Di Stasi LC. Anti-ulcerogenic and analgesic activities oa the leaves of
346 *Wilbrandia ebracteata* in mice. *Phytomedicine.* 2002; 9: 125-134.
- 347 34. Aguwa CN, Okunji CO. Gastrointestinal studies of *Pyrenacantha staudtii* leaf
348 extracts. *J Ethnopharmacol.* 1986; 15(1): 45-55.

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